

SSR ANALYSIS OF SOME *VITIS SYLVESTRIS* (GMEL.) ACCESSIONS OF THE SZIGETKÖZ AND FERTŐ-HANSÁG NATIONAL PARK, HUNGARY

G. JAHNKE^{1*}, Z. NAGY¹, G. KOLTAI², J. MÁJER¹

¹ National Agricultural Research and Innovation Centre, Research Institute for Viticulture and Enology, Badacsonytomaj, Hungary

² University of West Hungary Faculty of Agricultural and Food Sciences, Mosonmagyaróvár, Hungary

Abstract

The evolution of cultivated plants played important role in the ascent of humanity. Research of their origin and evolution started at the beginning of the 20th century, but till nowadays a lot of questions remain open. A large number of theories exist about the evolution of the European grapevine (*Vitis vinifera* L.). The *Vitis sylvestris* GMEL. in Hungary is a protected species. The quest and reservation of its populations are significant in terms of nature conservation and reserve of biodiversity as well. Based on theoretical and practical researches, it is supposed, that this species itself, or crossing with other species could be the progenitor of the European grapevine (*Vitis vinifera* L.).

In this study the quest and the SSR analysis of the *Vitis sylvestris* GMEL. populations of the Szigetköz and Fertő-Hanság National Park of Hungary are intended. 20 different genotypes of woodland grape (*Vitis sylvestris* GMEL.), 10 cultivars of European grape (*Vitis vinifera* L.) and 10 species/genotypes of rootstocks were analysed in 16 SSR loci of different linking groups.

The results show, that the analysed *Vitis sylvestris* accessions form a distinct group, but are closer to the *Vitis vinifera* cultivars, than to the rootstocks. This raises the probability, that these woodland grapes are true-to-type *Vitis sylvestris*.

Keywords: *Vitis sylvestris* (GMEL.), biodiversity, progenitor, SSR analysis, woodland grape, European grape, genotypes, true-to-type

1 INTRODUCTION

A large number of theories exist about the evolution of the European grapevine (*Vitis vinifera* L.). According to DE CANDOLLE (1894) the grape originate from the Trans-Caucasian part of Russia. Later the grape changed a lot under cultivation, and spread in larger and larger areas. After this, the wide-yielded grapes altered considerably under the effect of grown ones. In the after-glacial Eurasia, the existed woodland grape (*Vitis sylvestris* GMEL.) spread in whole Europe, and existed even in the southern part of Scandinavia. Once the *Vitis sylvestris* was taken into cultivation from the Trans-Caucasus by the peoples of the ancient Asia. Later the already *Vitis vinifera* was received by the peoples of the antique West-Asia and the people, who lived in the islands of the Aegean-see, spread it on the northern and southern bank of the Mediterranean-see (KOZMA, 1991).

According to TERPÓ (1986) the *Vitis vinifera* L. is not uniform, but the progeny of more original grape species, the main fundaments between the *Vitis sylvestris* GMEL. could be the hermaphrodite flowered *V. hissarica* and the *V. nuristanica*. In 1988 he developed a new intraspecific system of *Vitis sylvestris* GMEL. The substance of his taxonomy was that he sorted the woodland grapes into subspecies based on the hairs of the leaves, and into varietas based on the shape of the leaves. He deduced the eco-geographical groups (convarietas) of *Vitis vinifera* L. directly or indirectly from these varieties.

According to KOZMA (1991) the natural evolution produced the *Vitis sylvestris* GMEL., and the *Vitis vinifera* L. developed from this, due to deliberate cultivation. The most of the *Vitis* species are unisexual, and dioecious. The hermaphrodite-flowered form of *Vitis sylvestris* GMEL. was developed in the male-flowered individuals by bud mutation.

The microsatellite analysis of the grape can be traced back to the early nineties. A lot of SSR loci were identified and characterised in grapes (THOMAS and SCOTT, 1993; SCOTT et al., 2000; etc.) and were mapped (ADAM-BLONDON et al., 2004.; CONSTANTINI et al., 2007.).

The characterisation of grapevine cultivars by microsatellite DNA markers in Europe in the framework of an international cooperation called GENRES 081 (European Network for Grapevine Genetic Resources Conservation and Characterisation) was carried out between 1997 and 2002. In this research programme 6 microsatellite primer pairs were determined and suggested for the characterisation of cultivars. An European project The GRAPEGEN06 which can be considered as the continuation of the GENRES 081 project started in January 2007. The main objective of GrapeGen06 was to contribute to the successful long term preservation of the *Vitis* genetic resources for the use of future generations.

The *Vitis sylvestris* GMEL. in Hungary is a protected species. The quest and reservation of its populations are significant in terms of nature conservation and reserve of biodiversity as well.

A new project started in September 2013 in Hungary aimed in the quest and ex situ conservation of *Vitis sylvestris* individuals in the area of Szigetköz and Fertő-Hanság National Park as well, as the morphological description and analyses of them by molecular markers. The results of the planned analyses can go a long way into the clarification of the origin of *Vitis vinifera* L., and to the explanation of the development of the convarietases of the European grapevine.

2 MATERIALS AND METHODS

Plant material

The plant material (dormant canes) of 10 *Vitis* rootstocks, 10 *Vitis vinifera* L. varieties and 20 *Vitis sylvestris* accessions (listed in Table 1) for DNA extraction originated from the collection of the NAIRC RIVE in Badacsony (Hungary) and from the collection of the University of Pannonia in Cserszegtomaj (Hungary). The *V. sylvestris* materials from the Szigetköz and Fertő-Hanság National Park were reserved in Badacsony in 2010, and samples were collected in 2013.

No	ID/Label	Accession Name	Genetic Origin	Origin of the Accession
1	S1	Sylvestris B1	<i>Vitis sylvestris</i> GMEL.	Badacsony, Hungary (Collected from Szigetköz and Fertő-Hanság National Park)
2	S2	Sylvestris B2	<i>Vitis sylvestris</i> GMEL.	
3	S3	Sylvestris B3	<i>Vitis sylvestris</i> GMEL.	
4	S4	Sylvestris B4	<i>Vitis sylvestris</i> GMEL.	
5	S5	Sylvestris B5	<i>Vitis sylvestris</i> GMEL.	
6	S6	Sylvestris B6	<i>Vitis sylvestris</i> GMEL.	
7	S7	Sylvestris B7	<i>Vitis sylvestris</i> GMEL.	
8	S8	Sylvestris B8	<i>Vitis sylvestris</i> GMEL.	
9	S9	Sylvestris B9	<i>Vitis sylvestris</i> GMEL.	
10	S10	Sylvestris B10	<i>Vitis sylvestris</i> GMEL.	
11	S11	Sylvestris B11	<i>Vitis sylvestris</i> GMEL.	
12	S12	Sylvestris B12	<i>Vitis sylvestris</i> GMEL.	
13	S13	Sylvestris B13	<i>Vitis sylvestris</i> GMEL.	
14	S14	Sylvestris B14	<i>Vitis sylvestris</i> GMEL.	
15	S15	Sylvestris B15	<i>Vitis sylvestris</i> GMEL.	
16	S16	Sylvestris B16	<i>Vitis sylvestris</i> GMEL.	
17	S17	Sylvestris B17	<i>Vitis sylvestris</i> GMEL.	
18	S18	Sylvestris B18	<i>Vitis sylvestris</i> GMEL.	
19	ALHAROS	Alháros	<i>Vitis sylvestris</i> GMEL.	Cserszegtomaj, Hungary
20	DORGO	Dorgó	<i>Vitis sylvestris</i> GMEL.	
21	Sarga_ortlibi	Sárga ortlibi	<i>Vitis vinifera</i> L.	Badacsony, Hungary
22	Purcsin	Purcsin	<i>Vitis vinifera</i> L.	
23	Budai_goher	Budai góhér	<i>Vitis vinifera</i> L.	
24	Torok_goher	Török góhér	<i>Vitis vinifera</i> L.	
25	Balafant	Balafánt	<i>Vitis vinifera</i> L.	
26	Valtozo_Furmint	Változó Furmint	<i>Vitis vinifera</i> L.	
27	Piros_Furmint	Piros furmint	<i>Vitis vinifera</i> L.	
28	Koverszolo	Kövérzsölő	<i>Vitis vinifera</i> L.	
29	Primitivo	Primitivo	<i>Vitis vinifera</i> L.	
30	Fiano	Fiano	<i>Vitis vinifera</i> L.	
31	N._Mex.	V. Novo Mexicana	<i>V. riparia</i> x <i>V. candicans</i>	Cserszegtomaj, Hungary
32	Aramon_rup_G1	Aramon Ganzin N1	<i>V. vinifera</i> x <i>V. rupestris</i>	
33	V._vip._Ggb	Riparia Grand glabre	<i>V. riparia</i>	
34	V._rip._GdM	Gloire de Montpellier	<i>V. riparia</i>	
35	V._rup._FW1	Fort Worth N1	<i>V. rupestris</i>	
36	V._rup._FW2	Fort Worth N2	<i>V. rupestris</i>	
37	V._rup._FW3	Fort Worth N3	<i>V. rupestris</i>	
38	T5C	Teleki 5C E20	<i>V. berlandieri</i> x <i>V. riparia</i>	
39	SO4	Teleki-Fuhr SO4 (133)	<i>V. berlandieri</i> x <i>V. riparia</i>	
40	5BB	Teleki-Kober 5BB	<i>V. berlandieri</i> x <i>V. riparia</i>	

Table 1. List of the analyzed accessions

DNA extraction

DNA was extracted from the phloem of the dormant canes with DNA Plant Mini Kit (Quiagen), following the manufacturer's instructions. The amount and quality of DNA was determined spectrophotometrically. The DNA was diluted to a concentration of 10 ng/ml.

SSR analysis

Microsatellite (SSR) analysis was performed in 16 loci (Table 2). The primers had been chosen from different chromosomes (COSTANTINI et al., 2007) to give well defined heterozygosity.

Polymerase chain reactions were carried out in a total volume of 25 µl containing 12,5 µl of Hot Start Master Mix (Quiagen), 0.2 µM of each primer, and 50 ng of template DNA, using the following thermal profile: (1) 94°C for 45 min; (2) 94°C for 1 min, at the annealing temperature (Table 3) for 1 min, 73°C for 1 min per 35 cycles; (3) 73°C for 7 min.

One primer of each primer pairs were fluorescently labeled with FAM (6FAM) on the 5' end of the DNA chain. PCR products were run on a PE-Applied Biosystem 3100 Automated Capillary DNA Sequencer, the length of the products were determined using GeneScan 2.0 software (Applied Biosystem).

Linkage group ^a	SSR locus name	annealing temperature	Linkage group ^a	SSR locus name	annealing temperature
1	VMC8A7	64 °C	2	VMC7G3	60 °C
3	VVMD28	62 °C	4	VrZag21	62 °C
5	VrZag79	60 °C	8	VMC1F10	57 °C
9	VMC1C10	60 °C	10	VrZag25	67 °C
13	VMC3D12	57 °C	12	VMC2H4	57 °C
15	VMC5G8	58 °C	14	VMCNG1E1	58 °C
17	Scu06vv	60 °C	18	VVIM10	57 °C
19	VMC5E9	58 °C			

^aLinkage groups are numbered according to ADAM-BLONDON et al. (2004)

Table 2. List of the analyzed SSR loci.

Data analysis

Estimates of genetic similarity between pairs were calculated by the Jaccard index (JACCARD, 1908). For the generation of distance matrix and UPGMA dendogramm a demo version of MolMarker - a platform-independent software for the analyses of molecular marker data (under development by Gizella Jahnke and József Smidla) with fully and tested functionality in this issue - was used.

3 RESULTS AND DISCUSSION

The results of SSR analyses are shown in table 3. Based on these data a similarity matrix by Jaccard index was calculated (data not shown). Based on this matrix an UPGMA dendrogram was constructed (Figure 1.).

The main groups (*V. sylvestris*, *V. vinifera*, and rootstocks) mainly forms 4 distinct groups. It is visible, that the *Vitis sylvestris* GMEL. accessions — excluding Dorgó — form two distinct group in the dendrogram. The rootstock Aramon Ganzin N1 (*V. vinifera* x *V. rupestris*) takes place between the larger *V. sylvestris* group and the *V. vinifera* group, which is not surprising taking into account the hybrid origin of this accession. The larger *V. sylvestris* group is closer to the *Vitis vinifera* cultivars, than to the rootstocks. This raise the probability, that these woodland grapes are true-to-type *Vitis sylvestris*. The smaller *V. sylvestris* group, and Dorgó shows similarity to rootstocks, which probably shows their hybrid (*V. sylvestris* x *V. riparia*) origin.

To clarify these questions further genetic, molecular and morphological analyses are planned.

ID	VMC1F10	VMC3D12	VMC7G3	VVMD28	VMC2H4	VVIM10	VMC1C10	VrZag21	VMCNG1E1	VMC5G8	Scu06vv	VMC5E9	VMC4G6	VrZag79	VVS2	
S1	204	204	226	236	128	128	-1	-1	204	204	353	353	156	168	190	190
S2	204	204	206	236	-1	-1	-1	-1	-1	-1	156	168	190	190	93	101
S3	204	204	226	236	-1	-1	-1	-1	204	204	353	353	-1	-1	190	190
S4	204	204	226	236	-1	-1	-1	-1	204	204	-1	-1	-1	-1	190	190
S5	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	93	93
S6	204	204	-1	-1	-1	-1	-1	-1	200	200	-1	-1	-1	-1	194	194
S7	190	198	228	234	128	132	-1	-1	-1	-1	168	168	190	190	93	93
S8	202	204	-1	-1	-1	-1	250	250	-1	-1	327	327	-1	-1	-1	-1
S9	192	208	202	216	116	128	238	250	204	212	353	375	168	168	190	206
S10	-1	-1	228	228	130	132	222	222	200	204	353	365	156	168	190	194
S11	-1	-1	202	202	128	132	-1	-1	-1	-1	353	365	168	168	190	190
S12	190	190	202	202	116	128	-1	-1	204	204	353	365	156	156	190	200
S13	-1	-1	202	226	128	128	-1	-1	200	204	353	353	156	168	190	194
S14	190	208	226	234	128	132	238	238	200	204	353	365	156	168	190	194
S15	190	208	226	234	128	132	238	238	200	204	353	365	156	168	190	194
S16	190	208	226	234	128	132	238	238	200	204	353	365	156	168	190	194
S17	-1	-1	224	226	-1	-1	-1	-1	-1	-1	353	353	-1	-1	190	194
S18	-1	-1	224	226	-1	-1	-1	-1	-1	-1	353	353	-1	-1	190	194
ALHAROS	190	196	200	202	124	132	250	264	214	218	-1	-1	150	156	194	198
DORG	204	204	210	212	116	126	244	260	196	214	-1	-1	142	142	204	204
Sarga_ortlibi	190	196	222	238	-1	-1	230	238	200	224	353	353	-1	-1	200	206
Purcsin	196	204	216	216	-1	-1	-1	-1	198	220	353	365	-1	-1	202	206
Budai_goher	196	202	194	208	116	116	-1	-1	194	212	365	375	168	168	-1	-1
Torok_goher	-1	-1	216	222	116	116	-1	-1	194	212	365	373	-1	-1	105	105
Balafant	190	192	202	222	116	116	-1	-1	224	234	365	373	142	168	190	190
Valtozo_Furmint	192	204	216	222	116	116	230	250	206	236	365	365	142	168	200	206
Piros_Furmint	192	204	216	222	116	116	230	230	206	236	365	365	142	168	200	206
Koverszolo	192	204	216	222	116	116	230	230	206	236	365	365	142	168	200	206
Primitivo	202	204	224	226	-1	-1	250	260	200	206	365	365	-1	-1	200	206
Fiano	204	204	-1	-1	-1	-1	-1	-1	-1	-1	365	367	-1	-1	202	206
Novo_Mex.	194	200	198	198	122	130	248	248	194	202	365	365	142	150	204	210
Aramon_G1	192	198	198	214	118	118	238	262	212	236	363	363	156	168	200	214
G_g	194	196	196	196	122	126	234	250	200	202	363	363	142	142	206	212
G_d_M	196	196	196	206	-1	-1	-1	-1	204	212	363	363	142	142	204	210
FW_1	194	196	198	198	118	134	238	238	196	230	365	365	142	142	204	208
FW_2	194	202	198	224	122	134	238	248	-1	-1	363	363	142	142	204	208
FW_3	194	196	198	222	118	118	-1	-1	204	230	365	365	142	142	204	208
TSC	194	196	204	208	114	126	216	252	208	212	365	365	142	150	196	210
SO4	196	196	208	212	114	126	216	236	208	212	365	365	142	150	196	210
5BB	196	196	208	212	120	126	216	218	-1	-1	363	363	142	150	202	210

Table 3. Results of microsatellite (SSR) analysis in 15 loci. (-1 indicates missing data or null alleles)

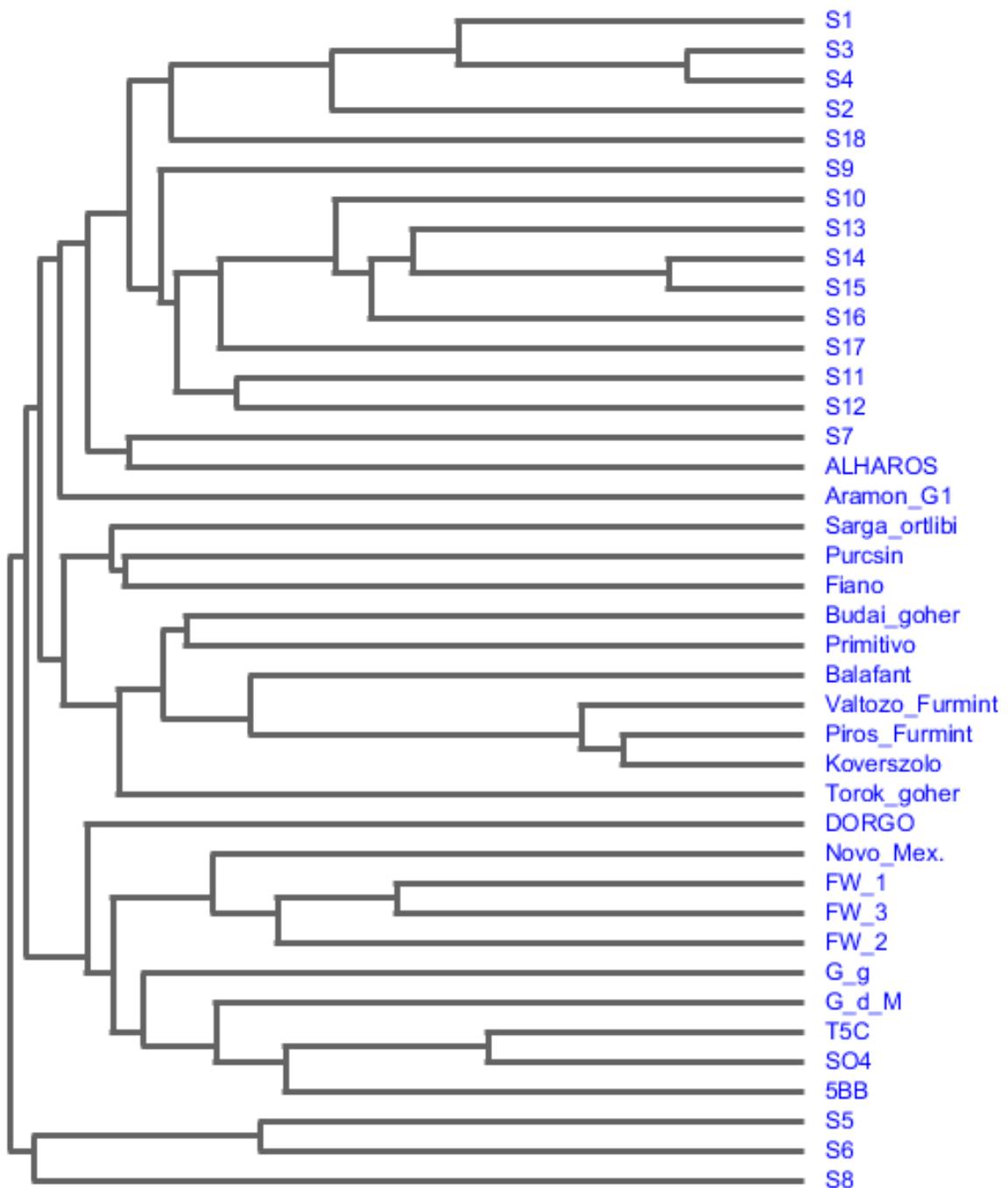


Figure 1. Dendogramm based on microsatellite results (Labels as in Table 1.).

4 ACKNOWLEDGEMENT

This research was funded by the Hungarian Scientific Research Fund (project no. PD-109386).

5 LITERATURE CITED

- ADAM-BLODON A. F., ROUX C., CLAUX D., BUTTERLIN G., MERDINOGLU D., THIS P. (2004): Mapping 245 SSR markers on the *Vitis vinifera* genome: a tool for grape genetics. *Theoretical and Applied Genetics* 109 : 1017-1027.
- COSTANTINI L., GRANDO M. S., FEINGOLD S., ULANOVSKY S., MEJIA N., HINRICHSEN P., DOLIGEZ A, THIS P., CABEZAS J. A., MARTINEZ-ZAPATER J. M. (2007): Generation of a Common Set of Mapping Markers to Assist Table Grape Breeding *American Journal of Enology and Viticulture* 2007 58: 102-111.
- DE CANDOLLE A. (1894): Termeszett növényeink eredete. 201- 204. p. Budapest: Királyi Magyar Természettudományi Társulat 516 p.
- KOZMA P. (1991): A szőlő és termesztése I. Budapest: Akadémiai Kiadó
- SCOTT K. D., LEE L. S., DOW T., HENRY R. J. (2000): Isolation and characterisation of new grape microsatellites. *Acta Horticulturae* 528: 199-200.
- TERPÓ A. (1986): A kultúrfajok eredete. 108-109. p. In: TERPÓ A (szerk.): Növényrendszertan az ökonómobotanika alapjaival I. Budapest: Mezőgazdasági Kiadó
- THOMAS M. R., SCOTT N. S. (1993): Microsatellite repeats in grapevine reveal DNA polymorphism when analysed as sequence tagged sites (STSs). *Theoretical and Applied Genetics* 86:895-990.